

# Dysbiosis of Gut Microbiome and Its Impact on Epigenetic Regulation

Eun-Sook Lee<sup>1</sup>,  
Eun-Ji Song<sup>1,2</sup> and  
Young-Do Nam<sup>1,2</sup>

## Abstract

High throughput methods have increased knowledge about the epigenome and microbiome, and allowed determination of the plausible link between the gut microbiome and epigenetic modification of the host. This has shed light on the development of various diseases such as immune-mediated, metabolic, and cardiovascular diseases, and even cancer. Dysbiosis, imbalanced gut microbiome which is frequently observed in such diseases, may be involved in regulating the epigenome of the host via direct changes in the gut microbiota or indirect changes of their metabolites, which are a variety of bioactive substances such as short chain fatty acids (SCFAs), biotin, folic acid, and other bioactive molecules. Indeed, correlation between host epigenetic regulation and alteration of gut microbiota or metabolites produced by intestinal microorganisms has been reported for various diseases. Therefore, the gut microbiome could be a diagnostic marker for certain diseases, and re-balancing dysbiosis through transplantation of the healthy gut microbiome could constitute an effective therapeutic strategy. Here, we discuss the relationship between dysbiosis of gut microbiota and the host epigenome, and suggest that the microbiome and epigenome are possible targets for disease diagnostics and therapeutics.

**Keywords:** Gut microbiota; Dysbiosis; Epigenome; Regulation

**Received:** April 14, 2017; **Accepted:** April 26, 2017; **Published:** May 02, 2017

## Introduction

Gene expression is regulated by epigenetic modifications such as DNA methylation, histone modifications, and binding of non-coding RNAs [1,2]. Gut microbiota can affect epigenetic processes of the host, consequently causing diseases such as allergy, Inflammatory Bowel Disease (IBD), autoimmune disease, metabolic syndrome, colorectal cancer, stress-related disorders, and neurodevelopmental disorders [3-9]. This possibly results from dysbiosis, a negative alteration of the gut microbiota, as well as changes in microbial metabolites, which are triggered by environmental factors such as diet, age, toxic chemicals, and pharmacological factors [10,11]. In fact, a pilot study reported a significant relationship between the predominant bacterial phyla and methylation patterns of the host in the human gut [12]. In addition, studies showed a strong influence of microbial metabolites such as Short-Chain Fatty Acids (SCFAs), in regulation of epigenetic programming in various tissues, including proximal colon, liver, and White Adipose Tissue (WAT) [13].

Few years ago, epigenome analysis, including analysis of DNA methylation and histone modifications, involved the use of

global or locus-specific methods such as High-Performance Capillary Electrophoresis (HPLC), Mass Spectrometry (MS), and western blot (11). However, recent developments in genome-wide high throughput methods such as Whole-Genome Bisulfite Sequencing (WGBS), reduced representation bisulfite sequencing (RRBS), methylated DNA immunoprecipitation sequencing (MeDIP-seq) for DNA methylation analysis, and Chromatin Immunoprecipitation (ChIP-seq) for histone modifications [11,14] has revolutionized epigenetic research. These methods have enabled determination of the relationship between the epigenome and the gut microbiome. Here, we will review the role of intestinal microorganisms in epigenetic regulation in terms of dysbiosis, the causative association of the gut microbiome with epigenetic modification of the host in diseases, and their potential for use in diagnostic and therapeutic strategies.

## Dysbiosis and Diseases

### Dysbiosis of gut microbiota

The diversity of the human gut microbiota has been extensively studied by high throughput analysis, revealing that the gut

- 1 Research Group of Gut Microbiome, Korea Food Research Institute, Sunnam, Republic of Korea
- 2 Department of Food Biotechnology, Korea University of Science and Technology, Daejeon, Republic of Korea

**Corresponding author:** Young-Do Nam

✉ youngdo98@kfri.re.kr

Research Group of Gut Microbiome, Korea Food Research Institute, Sunnam-si, Gyeonggi-do, Republic of Korea.

**Tel:** +82317809306

**Fax:** +82317099876

**Citation:** Lee ES, Song EJ, Nam YD. Dysbiosis of Gut Microbiome and Its Impact on Epigenetic Regulation. J Clin Epigenet. 2017, 3:S1.

microbiota varies between individuals but mainly consists of members of the Bacteroidetes and *Firmicutes* phyla [15,16]. Approximately one thousand different microorganisms co-inhabit the gut and they have crucial roles in maintaining homeostasis and health of the gastrointestinal tract [17,18]. Metchnikoff first coined the term “dysbiosis” [19] and the definition was extended to include a state in which changes in the diversity and abundance of gut microbiota, their metabolic activities, and local distribution produced harmful effects [20]. Actually, dysbiosis indicates an increase in the population of gut bacteria with pathogenic traits, which occasionally causes diseases [21,22]. For instance, an increase in the number of *Fusobacterium*, the pathogenic bacterium which was first detected in colon cancer, has been frequently observed in IBD [23]. Dysbiosis can be induced by various environmental factors such as diet, stress, and exposure to antibiotics, stress, toxins, drugs, and pathogens [22,24-26]. Accumulated evidence regarding clinical implications of dysbiosis via direct or indirect influences on related diseases, including immune disease and others, are reviewed below.

## Dysbiosis and diseases

Dysbiosis is closely related to systemic immune diseases including IBD, multiple sclerosis (MS) and autoimmune diseases [27-31]. Certain reports show that the change of symbiosis to dysbiosis in the intestinal tract is related to the development of systemic immune diseases through multiple routes such as changes in gut permeability and secretion of microbial enzymes [31,32]. The decrease in SCFAs during dysbiosis has significant implications in the regulation of the immune system, including intestinal barrier malfunction and reduction in anti-inflammatory effects [33,34]. In dysbiosis, the accumulated pathogenic bacteria increases intestinal barrier permeability and pathogens can be readily transferred to the host, causing diseases such as IBD [35,36]. In addition, protein-glutamine  $\gamma$ -glutamyltransferases (transglutaminases, Tgs) secreted by the gut microbiota under dysbiotic condition leads to altered posttranslational modification of the gut lumen, activating cascades of immune response that initiates pathological autoimmune processes [31].

In addition, some researchers have also examined the effects of dysbiosis on various other diseases. Jiang et al. recently reported that the dysbiosis of gut microbiota increased the permeability of the gut and the blood-brain barrier and mediated or activated pathogenesis of Alzheimer’s disease [37]. Tang et al. reviewed the possible molecular pathways that connect gut microbiota to cardiovascular or cardio metabolic diseases, such as the trimethylamine/trimethylamine N-oxide pathway, SCFA pathway, and the primary and secondary bile acids pathways [38]. Other studies showed that dysbiosis was related to obstructive sleep apnea-induced hypertension [39] and metabolic diseases such as obesity and impaired liver function [40,41].

## Dysbiosis and Epigenetic Regulation

### Direct relation between gut microbiota and epigenetic regulation

A study has demonstrated that gut microbiota can affect epigenetic regulation in immune homeostasis by direct

interaction with invariant natural killer cells (iNKTs) [42]. iNKT cells play important roles in ulcerative colitis (UC), IBD, and asthma by recognizing lipid antigens presented by CD1d molecules and secreting proinflammatory cytokines such as interleukin-4 (IL-4) and IL-13 [43,44]. Olszak et al. reported that microbial exposure to iNKT cells in germ-free mice decreased DNA methylation of the gene encoding the C-X-C motif chemokine ligand 16 (*Cxcl16*) and reduced its expression [42]. *Cxcl16* acts as a chemo-attractant of activated CD8 T cells, NKT cells, and Th1-polarized T cells [45], and therefore, reduced levels of *Cxcl16* leads to less accumulation of iNKT and improves barrier function. Therefore, the authors indicated the importance of contact with commensal microbes in early life for protection from immune-mediated disease such as IBD and asthma via restriction of the long-lasting activation of the iNKT pathway.

### Indirect relation between gut microbiota and epigenetic regulation through metabolites

Owing to the direct contact of diet-derived nutrients with the gut microbiome, nutritional modification can induce rapid shifts in the gut microbiome and probably cause diseases related to microbial dysbiosis [46]. Alterations in the gut microbiome may induce changes in microbial metabolites, which can be important but indirect regulators of the host epigenome. The intestinal microorganisms produce various low molecular weight bioactive substances including SCFAs, biotin, and other microbial metabolites such as folate and vitamin B<sub>2</sub>, which exert beneficial effects on physiological and epigenetic regulation [47,48].

### B-group vitamins produced by the gut microbiota

Most of the B vitamins such as riboflavin (vitamin B<sub>2</sub>), niacin (vitamin B<sub>3</sub>), pantothenic acid (vitamin B<sub>5</sub>), and biotin (vitamin B<sub>7</sub>) are synthesized by the gut microbiota [49]. Mammalian cells cannot synthesize B vitamins and thus, microbial production is partially required to meet the nutritional requirements of the host. In addition, the B vitamins produced by the gut microbiota contribute significantly to epigenomic processes [50]. For example, B vitamins act as cofactors of enzymes involved in epigenetic processes [51]. Nicotinamide adenine dinucleotide (NAD), the active form of niacin, is a cofactor of NAD-dependent histone deacetylases (HDACs), which catalyze the deacetylation of histones [52]. Pantothenate is the main acetyl group donor in the conversion of coenzyme A (Co-A) to acetyl-CoA [53]. Adequate supplies of acetyl-CoA support acetyl-CoA-dependent histone acetylation [54]. Biotinylation of histones [55] affects chromatin instability and thus cell proliferation, gene silencing, and the cellular response to DNA repair [56]. Folate acts as a key methyl group donor during the synthesis of precursors of nucleic acids and DNA methylation [57]. Folate is mainly supplied by the diet; however, it can also be synthesized by the colonic microbiota such as *Bifidobacterium bifidum* and *Bifidobacterium longum* [58,59]. Riboflavin (vitamin B<sub>2</sub>) serves as a cofactor of methylenetetrahydrofolate reductase (MTHFR), which is a folate-dependent enzyme involved in one-carbon metabolism, including DNA methylation [60]. Owing to its direct involvement

as a coenzyme in DNA methylation, the role of folate in epigenetic regulation is well-established. However, its role as a microbial metabolite in epigenetic regulation remains unknown [57]. According to Nagy-Szakal et al. supplementing pregnant mice with methyl donors such as folate, betaine, choline, and vitamin B<sub>12</sub> significantly changed microbiota composition in the offspring [61], indicating the putative role of microbiota-metabolized folate in epigenetic regulation.

### SCFAs

The most popular and well-studied microbial metabolites are SCFAs produced from indigestible carbohydrates as substrates of microbial fermentation. Indigestible carbohydrates include dietary fiber, resistant starch, oligosaccharides, and plant cell-wall polysaccharides [62]. The predominant SCFAs are acetate, butyrate, and propionate, which are produced mainly in the cecum and proximal colon in the molar ratio of 3:1:1 [63,64]. Theoretically, it is assumed that 10 g of indigestible carbohydrate fermentation yield around 100 mmol SCFAs [65]. Major groups of SCFA-producing gut microorganisms are listed in a review by Macfarlane et al. [62] SCFAs are absorbed in the cecum and proximal colon, from where they subsequently enter the mesenteric vein and drain into specific tissues; for instance, butyrate uptake is greater in the colonic epithelium, whereas propionate is preferentially utilized by the liver [66,67]. Thereafter, SCFAs contribute significantly to the daily energy requirement of the tissues and exert their physiological and epigenome-regulatory effects in various tissues [62].

Among SCFAs, butyrate and its source bacteria have been paid particular attention due to its multiple benefits, such as being the preferred energy source of colonic epithelial cells [68]. The most important butyrate-producing bacteria are *Faecalibacterium prausnitzii* (clostridial cluster IV), *Eubacterium rectale*, *Eubacterium hallii*, and *Roseburia* species (clostridial cluster XIVa) [68,69]. Its epigenetic function as a natural histone deacetylase inhibitor (HDACi) is well-established [70]. HDAC and histone acetyltransferases (HAT) are key enzymes that regulate gene expression by modulation of histone acetylation. Butyrate is a more potent natural HDACi that inhibits HDAC1/2 activity compared to acetate and propionate, which selectively inhibit HDAC2 and HDAC3, respectively [71-73]. This feature is utilized as a therapeutic strategy for treating diseases such as type 2 diabetes [74], cancer, and IBD [75,76].

### Dysbiosis and Epigenetic Regulation of Disease

Several studies regarding the putative effect of aberrant gut microbiome in epigenetic modulation of diseases exist. A recent pilot study showed a correlation between the predominant gut microbiota and differential DNA methylation of genes associated with the development of obesity and cardiovascular disease [12]. Furthermore, in their pilot birth cohort study, Tachibana et al. investigated whether the maternal gut microbiome influences the fetal risk of developing type 2 diabetes in the future by determining DNA methylation status using a high throughput

method, Infinium HumanMethylation450 BeadChip (Illumina), in 10 pregnant participants [77]. They identified an association between the proportion of Firmicutes in the maternal gut and the differential methylation rates in *UBE2E2* and *KCNQ1* in the umbilical cord samples, both of which are involved in insulin secretion. This study provided a possible connection between gut microbiota and epigenetic processes, particularly the methylation of type 2 diabetes-associated genes; however, further studies with a larger sample sizes are required. Dysbiosis of gut microbiota, characterized by higher numbers of the members of the genus *Bacteroides* and low butyrate production, is presumably induced by diet and can increase intestinal permeability followed by autoimmunity for type 1 diabetes [78]. This connection was explained by the differences in methylation of genes related to the pathogenesis of type 1 diabetes that were reported in their study.

Accumulated evidence demonstrated that SCFAs may act as intermediates in the interaction between the commensal microbiota and epigenetic modification of the host genome, resulting in diseases such as obesity, diabetes, and intestinal diseases such as ulcerative colitis (UC), Crohn's disease (CD), and colon cancer [79-83]. Schwiertz et al. reported differences in gut microbiota between lean and obese subjects and increased fecal SCFAs concentration in overweight individuals [82]. Furthermore, Remely et al. suggested that SCFA-producing bacteria and their products mediate the epigenetic regulation of gene expression [84]. In their study, lower diversity of the microbiota and scarcity of *Faecalibacterium prausnitzii* were observed in patients with obesity and type 2 diabetes compared to those in the lean control. The altered gut microbiota was concurrent with significantly lower methylation on the promoter region of SCFAs receptor *GPR41/FFAR3* in obese subjects.

Under microbial dysbiosis, a reduction or absence of butyrate-producing bacteria is a frequently observed feature in patients with UC and CD [85,86]. Nugent et al.'s metabolomic analysis in rectal biopsies of patients with colorectal adenomas showed a decreased butyrate level possibly due to the altered metabolome caused by microbial dysbiosis [87]. They suggested that these impairments are likely to contribute to the development of adenomas and colorectal cancer.

### Diagnostic and Therapeutic Application of the Role of Gut Microbiome on Epigenetic Regulation

Interest in the diagnostic and therapeutic application of the altered microbiome and epigenetic regulation in clinical practice is emerging. Determination of epigenomic modifications could be an effective approach for diagnosing and treating certain diseases [55]. Impaired DNA methylation, a predominant oncological feature, has been considered as a potential therapy target. Particularly, aberrant hypermethylation of specific CpG islands of tumor-suppressor genes, for instance, that of a BRCA1 carrier mutation, has been considered as good biomarkers for detecting the development of breast cancer [88]. Similarly,

assessment of microbial alteration also could be an effective diagnostic marker for certain diseases, including colon cancer and IBD [89-91]. Alterations in microbial community was identified at different states of colorectal carcinogenesis, with enrichment of microorganisms of the genus *Fusobacterium*, *Parvimonas*, *Gemella*, and *Leptotrichia*, and reduction in the numbers of *Bacteroides*, *Blautia*, *Sutterella*, *Collinsella aerofaciens*, and *Alistipes putredinis* in only the early stage of carcinogenesis. This implies that the identified bacteria could be candidates of a colorectal cancer-associated microbial marker [92]. Additionally, several reports regarding microbial dysbiosis frequently observed in early stage of IBD, suggesting a possible role of the gut microbiome as a diagnostic marker of IBD [93,94]. In addition, Maslowski et al. suggested the possibility of using SCFAs as a therapeutic agent as it can control inflammatory responses in immune disease such as colitis, arthritis, and asthma through the interaction with SCFA receptor, GPR43/FFAR2. Furthermore, Vrieze et al. investigated the effects of transferring gut microbiota from lean donors to patients with

metabolic syndrome and found an increase in insulin sensitivity and butyrate-producing microbiota in the feces, suggesting that gut microbiota can be used as therapeutic agents [95]. However, studies on the diagnosis and treatment of diseases using microbiome-epigenome correlation data are still in their infancy. This approach requires further research considering its cost-effectiveness and accuracy in application (Table 1).

## Conclusion

The human epigenome can be affected by various environmental factors, particularly those that affect the gut microbiota and their metabolites. The effects of the gut microbiome on host epigenetic regulation with respect to various diseases have been reviewed. Moreover, the link between the gut microbiome and the epigenome can be used as effective targets for the diagnosis and treatment of diseases. The recent developments in high-throughput technologies have broadened our understanding of gut microbiota and epigenomes, and it would serve as a key tool in identifying targets for diagnosis and treatment.

**Table 1:** Possible diagnosis and therapy using the interaction with gut microbiome and epigenetic.

Disease	Links to gut microbiome	Links to epigenetics	Diagnosis and therapy
Colorectal Cancer (CRC)	Increased abundance of <i>Fusobacterium nucleatum</i> and <i>Providencia</i> but decreased abundance <i>Lactobacillus</i> and butyrate-producing bacteria such as <i>Roseburia</i> and <i>Faecalibacterium</i> in CRC [96,97].	Butyrate, one of the most abundant SCFAs, is well known as HDACi which have antiangiogenic and antimetastatic effects in cancer [98,99], by epigenetically activating tumor-suppressor genes such as <i>p21</i> and <i>bax</i> [100] or suppressing carcinogenetic genes including <i>Cox-2</i> [101].	<ul style="list-style-type: none"> <li>• Oral administration of <i>Bifidobacterium</i> and <i>Bacteroides</i> which were suggested as therapeutic probiotics for cancer immunotherapy [102,103].</li> <li>• Butyrate [104] or the prebiotic sources such as acemannan in Aloe vera gel [105] with their chemopreventive effect.</li> </ul>
Diabetes	High ratio of Firmicutes/Bacteroidetes in type 2 diabetes with high abundance of lactic acid bacteria but low of <i>Faecalibacterium prausnitzii</i>	Changes in cell wall components such as LPS and peptidoglycan resulting from dysbiosis are involved in the epigenetic regulation of the inflammatory response [106-109].	<ul style="list-style-type: none"> <li>• Improved diets targeting to recover dysbiosis and epigenetic changes of pro-inflammatory genes in metabolic syndrome [106].</li> <li>• Transplantation of gut microbiota from lean and healthy donors to patients with metabolic syndrome (95).</li> </ul>
Obesity	Decreased production of butyrate by gut microbiota and lower diversity of the microbiota with low abundance of <i>F. prausnitzii</i> [84].	Hypomethylation at the promoter regions of SCFAs receptor GPR41/FFAR3 in obese patients.	<ul style="list-style-type: none"> <li>• GLP-1 agonist who contributed to the moderate increase of <i>F. prausnitzii</i> and the reverse of a hypomethylation of the promoter regions of GPR41/FFAR3 in patients with obesity and type 2 diabetes [84].</li> </ul>
IBD	Lower abundance of <i>Streptococcus</i> and the increased abundance of <i>Bacteroides</i> , <i>Parabacteroides</i> , and <i>Roseburia</i> [107]. Lower abundance of <i>Akkermansia muciniphila</i> in UC patients [108].	Hypomethylation at the differentially methylated regions (DMR) of KHDC3L (C6orf221) in UC patients, which were highly correlated with the dysbiosis [107]. Modulation of Fiaf, GPR43, HDACs, and PPAR $\gamma$ expression by <i>A. muciniphila</i> and propionate [83,109].	<ul style="list-style-type: none"> <li>• Identification of colonic mucosal DMRs can provide epigenetical and metagenomical targets for therapeutic measures [107].</li> <li>• Assessment of the gut microbial dysbiosis at the early stage of CD.</li> </ul>

## References

- 1 Cedar H, Bergman Y (2009) Linking DNA methylation and histone modification: patterns and paradigms. *Nat Rev Genet* 10: 295-304.
- 2 Jaenisch R, Bird A (2003) Epigenetic regulation of gene expression: how the genome integrates intrinsic and environmental signals. *Nat Genet* 33: 245-254.
- 3 Stilling RM, Dinan TG, Cryan JF (2014) Microbial genes, brain AND behaviour-epigenetic regulation of the gut-brain axis. *Genes Brain Behav* 13: 69-86.
- 4 Yang T, Owen JL, Lightfoot YL, Kladder MP, Mohamadzadeh M, et al. (2013) Microbiota impact on the epigenetic regulation of colorectal cancer. *Trends Mol Med* 19: 714-725.
- 5 Takahashi K, Sugi Y, Nakano K, Tsuda M, Kurihara K, et al. (2011) Epigenetic control of the host gene by commensal bacteria in large intestinal epithelial cells. *J Biol Chem* 286: 35755-35762.
- 6 Martinez-Medina M, Aldeguer X, Gonzalez-Huix F, Acero D, Garcia-Gil LJ (2006) Abnormal microbiota composition in the ileocolonic mucosa of Crohn's disease patients as revealed by polymerase chain reaction-denaturing gradient gel electrophoresis. *Inflamm Bowel Dis* 12: 1136-1145.
- 7 Frank DN, St Amand AL, Feldman RA, Boedeker EC, Harpaz N, et al. (2007) Molecular-phylogenetic characterization of microbial community imbalances in human inflammatory bowel diseases. *Proc Natl Acad Sci USA* 104: 13780-13785.
- 8 Wu HJ, Ivanov II, Darce J, Hattori K, Shima T, et al. (2010) Gut-residing segmented filamentous bacteria drive autoimmune arthritis via T helper 17 cells. *Immunity* 32: 815-827.
- 9 McLoughlin RM, Mills KH (2011) Influence of gastrointestinal commensal bacteria on the immune responses that mediate allergy and asthma. *J Allergy Clin Immunol* 127: 1097-107.
- 10 Lod S, Johansson T, Abrahamsson KH, Larsson L (2014) The influence of epigenetics in relation to oral health. *Int J Dent Hyg* 12: 48-54.
- 11 Torano EG, Garcia MG, Fernandez-Morera JL, Nino-Garcia P, Fernandez AF (2016) The Impact of External Factors on the Epigenome: In Utero and over Lifetime. *Biomed Res Int* 2016: s2568635.
- 12 Kumar H, Lund R, Laiho A, Lundelin K, Ley RE, Isolauri E, et al. (2014) Gut microbiota as an epigenetic regulator: pilot study based on whole-genome methylation analysis. *MBio*.
- 13 Krautkramer KA, Kreznar JH, Romano KA, Vivas EI, Barrett-Wilt GA et al. (2016) Diet-Microbiota Interactions Mediate Global Epigenetic Programming in Multiple Host Tissues. *Mol Cell* 64: 982-92.
- 14 Lara E, Calvanese V, Fernandez AF, Fraga MF (2011) Techniques to Study DNA Methylation and Histone Modification. Springer: London pp: 21-39.
- 15 Arumugam M, Raes J, Pelletier E, Le Paslier D, Yamada T, et al. (2011) Enterotypes of the human gut microbiome. *Nature* 473: 174-80.
- 16 Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, et al. (2005) Diversity of the human intestinal microbial flora. *Science* 308: 1635-1638.
- 17 Paracer S, Ahmadjian V (2000) Symbiosis: an introduction to biological associations (2<sup>nd</sup> edn.). Oxford University Press, NY, USA.
- 18 Eloe-Fadrosh EA, Rasko DA (2013) The human microbiome: from symbiosis to pathogenesis. *Ann Rev Med* 64: 145-163.
- 19 Murray MPJ (1988) Encyclopedia of natural medicine.
- 20 Hawrelak JA, Myers SP (2004) The causes of intestinal dysbiosis: a review. *Altern Med Rev* 9: 180-197.
- 21 Round JL, Mazmanian SK (2009) The gut microbiota shapes intestinal immune responses during health and disease. *Nat Rev Immunol* 9: 313-23.
- 22 Carding S, Verbeke K, Vipond DT, Corfe BM, Owen LJ (2015) Dysbiosis of the gut microbiota in disease. *Microb Ecol Health Dis* 26: 26191.
- 23 Strauss J, Kaplan GG, Beck PL, Rioux K, Panaccione R, et al. (2011) Invasive potential of gut mucosa-derived *Fusobacterium nucleatum* positively correlates with IBD status of the host. *Inflamm Bowel Dis* 17: 1971-1978.
- 24 Brown K, DeCoffe D, Molcan E, Gibson DL (2012) Diet-induced dysbiosis of the intestinal microbiota and the effects on immunity and disease. *Nutrients* 4: 1095-119.
- 25 Tanaka S, Kobayashi T, Songjinda P, Tateyama A, Tsubouchi M, et al. (2009) Influence of antibiotic exposure in the early postnatal period on the development of intestinal microbiota. *FEMS Immunol Med Microbiol* 56: 80-87.
- 26 O'Mahony SM, Marchesi JR, Scully P, Codling C, Ceolho AM, et al. (2009) Early life stress alters behavior, immunity, and microbiota in rats: implications for irritable bowel syndrome and psychiatric illnesses. *Biol Psychiatry* 65: 263-7.
- 27 Nagao-Kitamoto H, Kamada N (2017) Host-microbial Cross-talk in Inflammatory Bowel Disease. *Immune Netw* 17: 1-12.
- 28 Jia H, Hanate M, Aw W, Itoh H, Saito K, et al. (2017) Eggshell membrane powder ameliorates intestinal inflammation by facilitating the restitution of epithelial injury and alleviating microbial dysbiosis. *Sci Rep* 7: 43993.
- 29 Carrillo-Salinas FJ, Mestre L, Mecha M, Feliu A, Del Campo R, et al. (2017) Gut dysbiosis and neuroimmune responses to brain infection with Theiler's murine encephalomyelitis virus. *Sci Rep* 7: 44377.
- 30 Zechner EL (2017) Inflammatory disease caused by intestinal pathobionts. *Curr Opin Microbiol* 35: 64-69.
- 31 Lerner A, Aminov R, Matthias T (2017) Transglutaminases in Dysbiosis As Potential Environmental Drivers of Autoimmunity. *Front Microbio* 8: 66.
- 32 Honda K, Littman DR (2012) The microbiome in infectious disease and inflammation. *Annu Rev Immunol* 30: 759-95.
- 33 Fukuda S, Toh H, Hase K, Oshima K, Nakanishi Y, et al. (2011) Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature* 469: 543-547.
- 34 Ochoa-Reparaz J, Mielcarz DW, Ditrio LE, Burroughs AR, Foureau DM, et al. (2009) Role of gut commensal microflora in the development of experimental autoimmune encephalomyelitis. *J Immunol* 183: 6041-6050.
- 35 Michielan A, D'Inca R (2015) Intestinal Permeability in Inflammatory Bowel Disease: Pathogenesis, Clinical Evaluation, and Therapy of Leaky Gut. *Mediators Inflamm* 2015: 628157.
- 36 Vindigni SM, Zisman TL, Suskind DL, Damman CJ (2016) The intestinal microbiome, barrier function, and immune system in inflammatory bowel disease: a tripartite pathophysiological circuit with implications for new therapeutic directions. *Therap Adv Gastroenterol* 9: 606-25.
- 37 Jiang C, Li G, Huang P, Liu Z, Zhao B (2007) The Gut Microbiota and Alzheimer's Disease. *J Alzheimers Dis*.
- 38 Tang WH, Kitai T, Hazen SL (2017) Gut Microbiota in Cardiovascular Health and Disease. *Circ Res* 120: 1183-1196.

- 39 Durgan DJ (2017) Obstructive Sleep Apnea-Induced Hypertension: Role of the Gut Microbiota. *Curr Hypertens Rep* 19: 35.
- 40 Nicolas S, Blasco-Baque V, Fournel A, Gilleron J, Klopp P, et al. (2017) Transfer of dysbiotic gut microbiota has beneficial effects on host liver metabolism. *Mol Syst Biol* 13: 921.
- 41 Sen T, Cawthon CR, Ihde BT, Hajnal A, DiLorenzo PM, et al. (2017) Diet-driven microbiota dysbiosis is associated with vagal remodeling and obesity. *Physiol Behav* 173: 305-317.
- 42 Olszak T, An D, Zeissig S, Vera MP, Richter J, et al. (2012) Microbial exposure during early life has persistent effects on natural killer T cell function. *Science* 336: 489-493.
- 43 Fuss IJ, Heller F, Boirivant M, Leon F, Yoshida M, et al. (2004) Nonclassical CD1d-restricted NK T cells that produce IL-13 characterize an atypical Th2 response in ulcerative colitis. *J Clin Invest* 114: 1490-1497.
- 44 Cohen NR, Garg S, Brenner MB (2009) Antigen Presentation by CD1 Lipids, T Cells, and NKT Cells in Microbial Immunity. *Adv Immunol* 102: 1-94.
- 45 Matloubian M, David A, Engel S, Ryan JE, Cyster JG (2000) A transmembrane CXC chemokine is a ligand for HIV-coreceptor CXCR4. *Nat Immunol* 1: 298-304.
- 46 David LA, Maurice CF, Carmody RN, Gootenberg DB, Button JE, et al. (2014) Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 505: 559-563.
- 47 Jeffery IB, O'Toole PW (2013) Diet-microbiota interactions and their implications for healthy living. *Nutrients* 5: 234-252.
- 48 Paul B, Barnes S, Demark-Wahnefried W, Morrow C, Salvador C, et al. (2015) Influences of diet and the gut microbiome on epigenetic modulation in cancer and other diseases. *Clin Epig* 7: 112.
- 49 LeBlanc JG, Milani C, de Giori GS, Sesma F, van Sinderen D, et al. (2013) Bacteria as vitamin suppliers to their host: a gut microbiota perspective. *Curr Opin Biotechnol* 24: 160-168.
- 50 Noda H, Akasaka N, Ohsugi M (1994) Biotin Production by Bifidobacteria. *J Nutr Sci Vitaminol (Tokyo)* 40: 181-188.
- 51 Shenderov BA (2012) Gut indigenous microbiota and epigenetics. *Microb Ecol Health Dis*.
- 52 Penberthy WT, Kirkland JB (2012) Present Knowledge in Nutrition: Wiley-Blackwell pp: 293-306.
- 53 Siudeja K, Srinivasan B, Xu L, Rana A, de Jong J, et al. (2011) Impaired Coenzyme A metabolism affects histone and tubulin acetylation in Drosophila and human cell models of pantothenate kinase associated neurodegeneration. *EMBO Mol Med* 3: 755.
- 54 Cluntun AA, Huang H, Dai L, Liu X, Zhao Y, et al. (2015) The rate of glycolysis quantitatively mediates specific histone acetylation sites. *Cancer Metab* 3: 10.
- 55 Shenderov BA (2013) Metabiotics: novel idea or natural development of probiotic conception. *Microb Ecol Health Dis*.
- 56 Zemleni J (2005) Uptake, localization, and noncarboxylase roles of biotin. *Annu Rev Nutr* 25: 175-196.
- 57 Crider KS, Yang TP, Berry RJ, Bailey LB (2012) Folate and DNA Methylation: A Review of Molecular Mechanisms and the Evidence for Folate's Role. *Adv Nutr Int Rev J* 3: 21-38.
- 58 Pompei A, Cordisco L, Amaretti A, Zanoni S, Matteuzzi D, et al. (2007) Folate Production by Bifidobacteria as a Potential Probiotic Property. *Appl Environ Microbiol* 73: 179-185.
- 59 Strozzi GP, Mogna L (2008) Quantification of folic acid in human feces after administration of Bifidobacterium probiotic strains. *J Clin Gastroenterol* 2: S179-S184.
- 60 Oommen AM, Griffin JB, Sarath G, Zemleni J (2005) Roles for nutrients in epigenetic events. *J Nutr Biochem* 16: 747-747.
- 61 Nagy-Szakal D, Ross MC, Dowd SE, Mir SA, Schaible TD, et al. (2012) Maternal micronutrients can modify colonic mucosal microbiota maturation in murine offspring. *Gut Microbes* 3: 426-433.
- 62 Macfarlane GT, Macfarlane S (2012) Bacteria, colonic fermentation, and gastrointestinal health. *J AOAC Int* 95: 50-60.
- 63 Schwiertz A, Taras D, Schafer K, Beijer S, Bos NA, et al. (2010) Microbiota and SCFA in lean and overweight healthy subjects. *Obesity (Silver Spring)* 18: 190-195.
- 64 Wong JM, de Souza R, Kendall CW, Emam A, Jenkins DJ (2006) Colonic health: fermentation and short chain fatty acids. *J Clin Gastroenterol* 40: 235-243.
- 65 Cummings JH, Macfarlane GT (1997) Colonic microflora: nutrition and health. *Nutrition (Burbank, Los Angeles County, Calif)* 13: 476-478.
- 66 Cummings JH, Pomare EW, Branch WJ, Naylor CP, Macfarlane GT (1987) Short chain fatty acids in human large intestine, portal, hepatic and venous blood. *Gut* 28: 1221-1227.
- 67 Bloemen JG, Venema K, van de Poll MC, Olde Damink SW, Buurman WA, et al. (2009) Short chain fatty acids exchange across the gut and liver in humans measured at surgery. *Clin Nutr (Edinburgh, Scotland)* 28: 657-661.
- 68 Morrison DJ, Preston T (2016) Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism. *Gut Microbes* 7: 189-200.
- 69 Riviere A, Selak M, Lantin D, Leroy F, De Vuyst L (2016) Bifidobacteria and Butyrate-Producing Colon Bacteria: Importance and Strategies for Their Stimulation in the Human Gut. *Front Microbiol* 7: 979.
- 70 Ye J (2013) Improving insulin sensitivity with HDAC inhibitor. *Diabetes* 62: 685-687.
- 71 Cousens LS, Gallwitz D, Alberts BM (1979) Different accessibilities in chromatin to histone acetylase. *J Biol Chem* 254: 1716-1723.
- 72 Soliman ML, Rosenberger TA (2011) Acetate supplementation increases brain histone acetylation and inhibits histone deacetylase activity and expression. *Mol Cell Biochem* 352: 173-180.
- 73 Soliman ML, Smith MD, Houdek HM, Rosenberger TA (2012) Acetate supplementation modulates brain histone acetylation and decreases interleukin-1 $\beta$  expression in a rat model of neuroinflammation. *J Neuroinflammation* 9: 51.
- 74 Wang J, Tang H, Zhang C, Zhao Y, Derrien M, et al. (2015) Modulation of gut microbiota during probiotic-mediated attenuation of metabolic syndrome in high fat diet-fed mice. *ISME J* 9: 1-15.
- 75 Fofanova TY, Petrosino JF, Kellermayer R (2016) Microbiome-Epigenome Interactions and the Environmental Origins of Inflammatory Bowel Diseases. *J Pediatr Gastroenterol Nutr* 62: 208-219.
- 76 Berni Canani R, Di Costanzo M, Leone L (2012) The epigenetic effects of butyrate: potential therapeutic implications for clinical practice. *Clin Epigenet* 4: 4.
- 77 Tachibana K, Sakurai K, Watanabe M, Miyaso H, Mori C (2016) Associations between changes in the maternal gut microbiome and differentially methylated regions of diabetes-associated genes in fetuses: A pilot study from a birth cohort study. *J Diabetes Investig*.

- 78 Davis-Richardson AG, Triplett EW (2015) A model for the role of gut bacteria in the development of autoimmunity for type 1 diabetes. *Diabetologia* 58: 1386-1393.
- 79 Fofanova TY, Petrosino JF, Keller Mayer R (2016) Microbiome–Epigenome Interactions and the Environmental Origins of Inflammatory Bowel Diseases. *J Pediatr Gastroenterol Nutr* 62: 208-219.
- 80 Puddu A, Sanguineti R, Montecucco F, Viviani GL (2014) Evidence for the gut microbiota short-chain fatty acids as key pathophysiological molecules improving diabetes. *Mediators of Inflammation* 2014: 162021.
- 81 Duranton B, Keith G, Goss, Bergmann C, Schleiffer R, et al. (1998) Concomitant changes in polyamine pools and DNA methylation during growth inhibition of human colonic cancer cells. *Exp Cell Res* 243: 319-325.
- 82 Schwartz A (2009) Microbiota and SCFA in lean and overweight healthy subjects. *Obesity* 187: 190-195.
- 83 Maslowski KM, Vieira AT, Ng A, Kranich J, Sierro F, et al. (2009) Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 461: 1282-1286.
- 84 Remely M, Aumueller E, Merold C, Dworzak S, Hippe B, et al. (2014) Effects of short chain fatty acid producing bacteria on epigenetic regulation of FFAR3 in type 2 diabetes and obesity. *Gene* 537: 85-92.
- 85 Louis P, Flint HJ (2009) Diversity, metabolism and microbial ecology of butyrate-producing bacteria from the human large intestine. *FEMS Microbiol Lett* 294: 1-8.
- 86 Wang W, Chen L, Zhou R, Wang X, Song L, et al. (2014) Increased proportions of Bifidobacterium and the Lactobacillus group and loss of butyrate-producing bacteria in inflammatory bowel disease. *J Clin Microbiol* 52: 398-406.
- 87 Nugent JL, McCoy AN, Addamo CJ, Jia W, Sandler RS, et al. (2014) Altered tissue metabolites correlate with microbial dysbiosis in colorectal adenomas. *J Proteome Res* 13: 1921-1929.
- 88 Rykova EY, Tsvetovskaya GA, Sergeeva GI, Vlassov VV, Laktionov PP (2008) Methylation-based analysis of circulating DNA for breast tumor screening. *Ann N Y Acad Sci* 1137: 232-235.
- 89 Da Silva K (2014) Microbiota: Dysbiosis as a diagnostic. *Nat Med* 20: 348.
- 90 Gagniere J, Raisch J, Veziat J, Barnich N, Bonnet R, et al. (2016) Gut microbiota imbalance and colorectal cancer. *World J Gastroenterol* 22: 501-518.
- 91 Dickson I (2017) Gut microbiota: Diagnosing IBD with the gut microbiome. *Nat Rev Gastroenterol Hepatol* 14: 195.
- 92 Nakatsu G, Li X, Zhou H, Sheng J, Wong SH, Wu WK, et al. (2015) Gut mucosal microbiome across stages of colorectal carcinogenesis. *Nat Commun* 6: 8727
- 93 Gevers D, Kugathasan S, Knights D, Kostic AD, Knight R, et al. (2017) A Microbiome Foundation for the Study of Crohn's Disease. *Cell Host Microbe* 21: 301-304.
- 94 Gevers D, Kugathasan S, Denson LA, Vazquez-Baeza Y, Van Treuren W, et al. (2014) The treatment-naïve microbiome in new-onset Crohn's disease. *Cell Host Microbe* 15: 382-392.
- 95 Vrieze A, Van Nood E, Holleman F, Salojarvi J, Kootte RS, et al. (2012) Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology* 143: 913-6.e7.
- 96 Wu N, Yang X, Zhang R, Li J, Xiao X, Hu Y, et al. (2013) Dysbiosis signature of fecal microbiota in colorectal cancer patients. *Microb Ecol* 66: 462-470.
- 97 Burns MB, Lynch J, Starr TK, Knights D, Blehman R (2015) Virulence genes are a signature of the microbiome in the colorectal tumor microenvironment. *Genome Med* 7: 55
- 98 Bolden JE, Peart MJ, Johnstone RW (2006) Anticancer activities of histone deacetylase inhibitors. *Nat Rev Drug Discov* 5: 769-784.
- 99 Liang D, Kong X, Sang N (2006) Effects of histone deacetylase inhibitors on HIF-1. *Cell Cycle* 5: 2430-2435.
- 100 Dashwood RH, Ho E (2007) Dietary histone deacetylase inhibitors: from cells to mice to man. *Semin Cancer Biol* 17: 363-369.
- 101 Tong X, Yin L, Giardina C (2004) Butyrate suppresses Cox-2 activation in colon cancer cells through HDAC inhibition. *Biochem Biophys Res Commun* 317: 463-471.
- 102 Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, et al. (2015) Commensal Bifidobacterium promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science* 350: 1084-1089.
- 103 Vetizou M, Pitt JM, Daillere R, Lepage P, Waldschmitt N, et al. (2015) Anticancer immunotherapy by CTLA-4 blockade relies on the gut microbiota. *Science* 350: 1079-1084.
- 104 Crim KC, Sanders LM, Hong MY, Taddeo SS, Turner ND, et al. (2008) Upregulation of p21Waf1/Cip1 expression in vivo by butyrate administration can be chemoprotective or chemopromotive depending on the lipid component of the diet. *Carcinogenesis* 29: 1415-1420.
- 105 Al-Madbolly LA, Kabbash A, Yassin AM, YAGI A (2017) Dietary cancer prevention with butyrate fermented by Aloe vera gel endophytic microbiota. *J Gastroenterol Hepatol* 6: 2312-2317.
- 106 Remely M, Aumueller E, Jahn D, Hippe B, Brath H, Haslberger AG (2015) Microbiota and epigenetic regulation of inflammatory mediators in type 2 diabetes and obesity. *Beneficial Microbes* 5: 33-43.
- 107 Harris RA, Shah R, Hollister EB, Tronstad RR, Hovdenak N, et al. (2016) Colonic Mucosal Epigenome and Microbiome Development in Children and Adolescents. *J Immunol Res* 2016: 9170162.
- 108 Shah R, Cope JL, Nagy-Szakal D, Dowd S, Versalovic J, et al. (2016) Composition and function of the pediatric colonic mucosal microbiome in untreated patients with ulcerative colitis. *Gut Microbes* 7: 384-396.
- 109 Lukovac S, Belzer C, Pellis L, Keijsers BJ, de Vos WM, et al. (2014) Differential modulation by Akkermansia muciniphila and Faecalibacterium prausnitzii of host peripheral lipid metabolism and histone acetylation in mouse gut organoids. *mBio*.